

Full Length Research Paper

Field performance of *shrunk-2* super-sweet corn populations derived from tropical field maize × *shrunk-2* super-sweet corn crosses in Ibadan, Nigeria

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Received 26 December, 2018; Accepted 3 April, 2019

The conversion of tropical field corn genotypes into sweet corn could broaden the genetic base and improve yield and adaptation of sweet corn varieties. In this study, the performance of *shrunk-2* (*sh2*) super-sweet corn populations derived from crosses between a *sh2* population and tropical field corn genotypes were evaluated in Ibadan. Experiments were conducted using randomised complete block design with three replicates. Data were collected on agronomic and fresh ear yield traits, and then subjected to analysis of variance. Significant genotypic differences were observed among the populations with most of the derived populations significantly superior to the donor population for most of the traits. Yield of marketable cobs ranged from 5.80 to 7.63 t/ha (mean = 6.84 t/ha). Six derived populations had significantly higher yield of marketable cobs than the donor population. On the average, 83.1% of the number of cobs harvested was marketable. Husk cover scores ranged from 2.8 to 6.8, with all the derived populations having significantly lower husk cover scores than the donor population. The results indicated that the conversion of the field corn genotypes into super-sweet corn was effective in the development of new super-sweet corn populations. The observed genetic differences could be exploited in further breeding programmes.

Key words: Fresh ear yield, husk cover, *shrunk-2* super-sweet corn, yield of marketable cobs

INTRODUCTION

Maize (*Zea mays* L.) is an important and highly diversified crop in the world, and a cereal food staple cultivated in every agro-ecological zone in West and Central Africa. It is made up of different types, classified by their kernel endosperm characteristics. The most common types are; flint, dent, flour, pop, pod, waxy and sweet. In West and Central Africa, the flint and dent types are the most widely cultivated (Kim et al., 1987; Kim and Ajala, 1996). These field maize types, in addition to the

dry grains being consumed in different processed forms, are harvested and consumed fresh as green maize (after roasting, steaming or boiling on the cob). This contrasts with developed countries where human consumption of fresh maize is of sweet and super-sweet corn (Osayintola et al., 1992; Lee et al., 1999; Zan and Brewbaker, 1999). The widely grown field maize varieties in West and Central Africa were however not specifically developed for direct fresh human consumption (Ogunbodede, 1999).

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Sweet corn (*Zea mays* L. var *saccharata*), is a type of maize with kernels that are sweet as a result of high sugar content which when consumed in the fresh immature stage, has high levels of total sugars than field maize, rich in fibre, minerals, certain vitamins (Tracy, 1997; Lertrat and Pulam, 2007), and significant antioxidant properties (Dewanto et al., 2002). Sweet corn has its origin from a mutation that influences carbohydrate biosynthesis in the endosperm. In the genome of sweet corn, at least one of the eight mutant genes preventing the conversion of sugars to starch is present. These genes, which include *shrunk-2* (*sh2*) on chromosome 3; brittle (*bt*), and amylose extender (*ae*) on chromosome 5; sugary enhancer (*se*), sugary (*su*), and brittle-2 (*bt2*) on chromosome 4; dull (*du*) on chromosome 10; and waxy (*wx*) on chromosome 9 (Tracy et al. 2006, Qi et al., 2009) are monogenic and recessive (Santos et al., 2014). Of all the mutant types, *sh2* (Yousef and Juvik, 2002) and *bt2* (Brewbaker, 1977) have the greatest commercial value.

The *sh2* types of sweet corn contain about 29.9% sucrose, which is about ten times that of field maize. These types of sweet corn, generally referred to as super-sweet or extra-sweet corn, have extended shelf life due to the slow conversion of sugars to starch after harvest (Tracy, 1997), but with highly reduced levels of total carbohydrate. The *sh2* gene reduces endosperm content of starch and water soluble polysaccharides which leads to a reduction in the energy content of maize kernels and the markedly collapsed physical appearance of the kernels when dry. Due to these features, *sh2* varieties generally have a significantly reduced germination, seedling emergence, seedling vigour, plant development and growth and poor stand establishment.

In Nigeria, maize is grown throughout the country from the high rainfall forest of the southeast to the low rainfall Sudan savanna of the north; and with supplemental irrigation, maize can be grown throughout the year. Nigeria produces about 40% of the maize production in West and Central Africa (FAO, 2016). Great potential therefore exists for the production of super-sweet corn in Nigeria. Sweet corn production is usually targeted at three distinct and largely independent markets, namely; fresh, canning and freezing, with the fresh market component accounting for more than 70% of the total (Lizaso et al., 2007; USDA, 2017). Imports of canned sweet corn to Nigeria have increased in recent years largely due to widening food preferences. Therefore, the development and production of super-sweet corn in large quantity and quality would impact positively on the social and economic life of Nigeria. However, sweet corn cultivars are virtually nonexistent in Nigeria, and coupled with poor adaptation to tropical environments, its cultivation and utility is limited.

To bridge the gap created by these challenges, a broad-base temperate super-sweet *sh2* corn population was introduced into the country and adapted to the

prevailing tropical environmental conditions by four cycles of mass selection (Adetimirin, 2008). This was meant to serve both as an open pollinated variety, as well as basis for inbred line development and hybrid production. The performance of this *sh2* super-sweet population in the growing conditions of Nigeria could be improved. One way of achieving this is by converting tropical field maize genotypes into sweet corn through backcrossing and selection for the sweet corn trait. This strategy will lead to improvement in the yield of the super-sweet corn varieties, broaden the narrow genetic base characteristic of *sh2* (Tracy, 2001; Teixeira et al., 2013), and facilitate the development of *sh2* inbred lines and hybrids. According to Entringer et al. (2017), the backcross method of breeding is efficient for obtaining super-sweet corn populations with good agronomic performance. Previous studies Carrea et al. (1996), Malvar et al. (1997, 2001), Tracy (2001), Butrón et al. (2008) and Entringer et al. (2017) reported the use of field corn to improve the agronomic performance of sweet corn. These studies have also shown that field maize genotypes could differ in their ability to improve the agronomic performance and quality of sweet corn. Therefore, tropical normal endosperm field maize genotypes have the potential to improve the adaptation and productivity of super-sweet corn. The objective of this study was to evaluate the agronomic performance of some *sh2* super-sweet corn populations derived from crosses between a tropicalised *sh2* population and tropical normal endosperm field corn genotypes.

MATERIALS AND METHODS

Location of experimental site

The study was conducted at the experimental field of the Department of Agronomy, Faculty of Agriculture, along Parry road, University of Ibadan (7°26' N, 3°54' E), Ibadan, Nigeria.

Generation of new homozygous *sh2* super-sweet corn populations

A broad-based temperate super-sweet *sh2* maize population, which was introduced and adapted to tropical environmental conditions of Ibadan, Nigeria after four cycles of mass selection (Adetimirin, 2008), was crossed as male (using bulk pollen) to six tropical field maize inbred lines (1368, 4001, 4008, 9613, KU1409 and KU1414) and five commercial/experimental hybrids [1368/9071 (Oba super 1), 4001/KU1414 (Oba super 2), KU1409/9613, KU1409/4008 and 4001/4008], all developed at the International Institute of Tropical Agriculture (IITA), Ibadan, Nigeria

The resulting F1 populations from each of these initial crosses were sib-mated to generate F2 populations. The F2 seeds were thereafter selected for kernels homozygous for the *sh2* phenotype. The selected homozygous *sh2* derived from each of the F2 populations were backcrossed to the recurrent (field maize) parents. The BC1 plants from each population were sib-mated to generate BC1S1 which were thereafter selected for kernels homozygous for the *sh2* phenotype. The selected homozygous *sh2* derived from each of the BC1S1 populations were then bulked to

Table 1. Codes of *sh2* populations and their corresponding field maize parent genotypes.

S/N	Code	Field maize parent
1	Pop1	KU1409
2	Pop2	1368
3	Pop3	KU1409/4008
4	Pop4	KU1414 SR
5	Pop5	KU1409/9613
6	Pop6	Oba Super 1
7	Pop7	4001/4008
8	Pop8	Oba Super 2
9	Pop9	4008
10	Pop10	9613
11	Pop11	4001
12	<i>sh2</i> donor Pop	

form new homozygous *sh2* populations as shown in Table 1.

Evaluation of *sh2* super-sweet corn populations

The eleven new homozygous derived *sh2* populations alongside the original donor *sh2* population were evaluated in replicated field trials under rain-fed conditions during the 2016 and 2017 cropping seasons. The soil at the experimental site is sandy-loam with 15.20 g kg⁻¹ organic carbon, 0.98 g kg⁻¹ total nitrogen, 14.79 mg kg⁻¹ available P (Bray-1), 0.26 cmol kg⁻¹K and a pH(H₂O) of 6.1. The twelve *sh2* populations were arranged in a randomized complete block design with three replicates. Each plot consisted of four 5.0 m long rows, with plants spaced 0.50 m and rows 0.75 m apart. Planting was done on the flat. Plots were over-sown to ensure good plant stand (since germination failure is a common feature in super-sweet corn with the *sh2* gene) and later thinned to two plants at two weeks after planting (WAP) to give a plant population of approximately 53,333 plants per hectare. NPK 15-15-15 fertilizer was applied at the rate of 300 kg ha⁻¹ at 2 WAP, which provided 45 kg N ha⁻¹, 20 kg P ha⁻¹ and 36 kg K ha⁻¹ and top-dressed using urea at the rate of 25 kg N ha⁻¹ at 4 WAP. Plots were kept weed free by hand weeding.

Data collection

Data were collected on days to anthesis (DA) as number of days from planting to when 50% of the plants in a plot shed pollen, and days to silking (DS) as number of days from planting to when 50% of the plants in a plot have emerged silks. Anthesis-silking interval (ASI) was calculated as the difference in days between DS and DA. Plant height (PH) and ear height (EH) were measured in meters 2 weeks after silking on all plants in the two middle rows of a plot, as the average distance from the soil level to the collar of the uppermost leaf and collar of the leaf bearing the uppermost ear, respectively. Husk cover (HC) was scored on plot basis on a scale of 1 to 9 (1 = husk tightly covers ear tip and extends beyond it; 9 = poor husk cover with ear tip clearly exposed). Harvesting for yield data was carried out between stages R4 and R5 when the ears were still green and fresh, 21 days after silking using plants in the two middle rows of a plot. Yield data included: (i) number of cobs (NC) recorded as total number of ears (fresh ears with husk removed) harvested per plot expressed per hectare; (ii) yield of

cobs (YC) recorded as total weight of cobs (fresh ears with husk removed) harvested per plot expressed in tonnes per hectare; (iii) number of marketable cobs (NMC) recorded as number of cobs with approximately 250 filled edible kernels per plot, expressed per hectare; (iv) yield of marketable cobs (YMC) recorded as total weight of marketable cobs per plot expressed in tonnes per hectare; (v) number of kernel rows (NKR) recorded as the average number of kernel rows of 10 top cobs; (vi) cob length (CL) measured in cm as the average length of 10 top cobs; (vii) cob diameter (CD) was measured using an electronic 6 in digital calliper (Pittsburgh®, Item #47257) as the average diameter of 10 top cobs taken at the middle portion of the cob.

Data analyses

All data were subjected to analysis of variance for a randomized complete block using the PROC. GLM procedure in SAS (SAS Institute, 2003) assuming a mixed model with genotype and the interaction between genotype and environment were considered random. The years were considered as separate environments. Significant means were separated using DMRT (p=0.05).

RESULTS

Significant genotype differences were observed for all traits, except ASI. Effect of years was only significant for YC, NC and EH. Significant interactions between genotypes and environment were also found for most of the traits except YC, NC, HC, EH and ASI. However, the contribution of the main effects of genotype to the total sum of squares was in all cases larger than that due environment or interaction of genotype and environment (Table 2). Mean performance of the 12 populations over two years is presented in Table 3. The range in YMC from the lowest value (5.80 t/ha) to the highest value (7.63 t/ha) was 26.7% of the mean YMC (6.84 t/ha). However, six of the derived populations had significantly higher YMC than the donor *sh2* composite population. The YMC obtained in the present study was 95.5% of YC.

Table 2. Mean squares from analysis of variance on the effects of environment and genotype on agronomic and fresh ear yield traits averaged across two years of 12 *shrunken-2* super-sweet corn populations evaluated across two years in Ibadan, Nigeria.

Source of variation	df	Mean square												
		YMC (t/ha)	NMC	YC (t/ha)	NC	NKR	CL (cm)	CD (mm)	HC	PH (cm)	EH(cm)	DA	DS	ASI
Rep(Env)	4	0.43*	6.73	0.30	4.36	0.11	0.08	1.48	0.31	52.66	90.11*	0.69**	0.51	0.61
Environment	1	1.31	0.39	2.47*	33.04*	0.59	0.59	5.39	0.01	277.30	475.35**	1.13	2.35	0.22
Genotype	11	2.56**	40.07**	3.65**	61.20**	2.94**	2.14*	10.69**	7.86**	230.57**	389.72**	6.59**	5.14**	1.65
Env*Genotype	11	0.59**	9.10**	0.36	4.06	0.39**	0.55**	2.24**	0.50	78.36**	33.86	0.43*	1.01**	1.07**
Error	44	0.16	3.12	0.25	3.63	0.12	0.13	0.72	0.35	27.62	30.69	0.18	0.33	0.34

YMC: Yield of marketable cobs; NMC: Number of marketable cobs; YC: Yield of cobs; NC: Number of cobs; NKR: Number of kernel rows; CL: Cob length; CD: Cob diameter; HC: Husk cover score; PH: Plant height; EH: Ear height; DA: Days to anthesis; DS: Days to silking; ASI: Anthesis-silking interval; *, **: significant respectively at 0.05, 0.01 probability levels.

Table 3. Genotypic means of agronomic and fresh ear yield traits averaged across two years of 12 *shrunken-2* super-sweet corn populations evaluated across two years in Ibadan, Nigeria.

Genotype	YMC (t/ha)	NMC ($\times 10^3$)	YC (t/ha)	NC ($\times 10^3$)	NKR	CL (cm)	CD (mm)	HC	PH (cm)	EH (cm)	DA	DS	ASI
Pop1	6.51 ^b	40.82 ^b	6.83 ^b	50.50 ^b	13.8 ^{ef}	15.0 ^e	44.3 ^{cd}	3.3 ^{ef}	2.05 ^{ab}	0.97 ^{cd}	55.8 ^{bc}	58.3 ^{abc}	2.5 ^{bcd}
Pop2	6.59 ^b	37.88 ^{cd}	6.94 ^b	46.38 ^{de}	15.3 ^{ab}	16.1 ^{bc}	47.3 ^a	5.8 ^b	2.07 ^{ab}	1.04 ^b	56.2 ^{abc}	58.7 ^{ab}	2.5 ^{bcd}
Pop3	7.21 ^a	43.73 ^a	7.56 ^a	53.33 ^a	13.0 ^h	15.4 ^{de}	43.2 ^d	3.7 ^{de}	1.88 ^f	0.86 ^e	55.7 ^c	58.7 ^{ab}	3.0 ^{abc}
Pop4	7.31 ^a	45.30 ^a	7.60 ^a	50.10 ^b	13.8 ^{ef}	15.4 ^{de}	45.9 ^b	3.3 ^{ef}	2.00 ^{bcd}	1.06 ^b	55.7 ^c	58.0 ^{bcd}	2.3 ^{cd}
Pop5	5.80 ^c	36.71 ^d	6.18 ^{cd}	43.93 ^f	14.7 ^{bc}	14.3 ^f	44.4 ^{cd}	4.0 ^{de}	2.00 ^{bcd}	1.03 ^{bc}	54.0 ^d	56.5 ^f	2.5 ^{bcd}
Pop6	7.43 ^a	39.29 ^{bc}	7.83 ^a	43.30 ^f	15.0 ^{ab}	15.8 ^{bcd}	46.9 ^{ab}	4.3 ^{cd}	1.95 ^{de}	0.94 ^d	54.0 ^d	56.3 ^f	2.3 ^{cd}
Pop7	7.63 ^a	41.55 ^b	8.11 ^a	49.86 ^{bc}	14.2 ^{de}	16.6 ^a	45.9 ^b	3.7 ^{de}	1.90 ^{ef}	0.97 ^{cd}	56.3 ^{ab}	57.8 ^{cd}	1.5 ^e
Pop8	7.53 ^a	40.63 ^b	7.79 ^a	44.45 ^{ef}	14.0 ^{def}	16.2 ^b	46.3 ^{ab}	3.7 ^{de}	2.06 ^{ab}	1.06 ^b	54.3 ^d	56.8 ^{ef}	2.5 ^{bcd}
Pop9	6.11 ^{bc}	37.76 ^{cd}	6.59 ^{bc}	49.42 ^{bc}	13.7 ^{fg}	15.9 ^{bcd}	43.8 ^{cd}	3.8 ^{de}	2.04 ^{abc}	1.08 ^b	56.0 ^{bc}	59.0 ^a	3.0 ^{abc}
Pop10	6.26 ^{bc}	38.14 ^{cd}	6.57 ^{bc}	47.55 ^{cd}	14.3 ^{cd}	15.6 ^{cd}	44.4 ^{cd}	2.8 ^f	2.03 ^{abc}	1.16 ^a	54.3 ^d	57.5 ^{de}	3.2 ^{abc}
Pop11	7.50 ^a	39.40 ^{bc}	8.16 ^a	51.50 ^{ab}	13.3 ^{gh}	16.1 ^{bc}	45.8 ^b	4.8 ^c	1.97 ^{cd}	0.93 ^{de}	56.7 ^a	58.8 ^a	2.2 ^{de}
<i>sh2</i> donor Pop	6.27 ^{bc}	38.16 ^{cd}	5.82 ^d	46.87 ^d	15.0 ^{ab}	15.6 ^d	44.0 ^{cd}	6.8 ^a	1.95 ^{de}	0.98 ^{cd}	53.8 ^d	57.3 ^{de}	3.5 ^a
Mean	6.84	39.95	7.16	48.1	14.2	15.7	45.2	4.2	1.99	1.01	55.2	57.8	2.6
CV (%)	5.91	4.42	6.98	3.96	2.47	2.30	1.88	14.17	2.64	5.51	0.77	1.00	22.52

YMC: Yield of marketable cobs; NMC: Number of marketable cobs; YC: Yield of cobs; NC: Number of cobs; NKR: Number of kernel rows; CL: Cob length; CD: Cob diameter; HC: Husk cover score; PH: Plant height; EH: Ear height; DA: Days to anthesis; DS: Days to silking; ASI: Anthesis-silking interval.

The YC ranged from 5.82 to 8.16 t/ha. The range in NC and NMC were 43,290 to 53,330 and

36,710 to 45,300 cobs/ha, respectively. On the average, 83.1% of the total NC harvested was

marketable. The NKR, CL and CD averaged 14.2, 15.7 cm and 45.2 mm, respectively.

One important quality attribute in fresh market sweet corn production is HC. In the present study, HC scores ranged from 2.8 to 6.8 with a mean of 4.2. All the derived *sh2* super-sweet corn populations had significantly lower HC scores than the donor *sh2* population, eight of which had scores lower than the mean. The PH and EH of the populations ranged from 1.9 to 2.1 m (mean = 2.0 m) and 0.9 to 1.2 m (mean = 1.0 m), respectively. On the average, ears were placed mid-way the height of the plants.

All the population including the *sh2* donor population exhibited intermediate maturity with DA and DS ranging from 53.8 to 56.7 days and 56.3 to 59.0 days, respectively. However, nine of the derived populations shed pollen significantly later than the donor *sh2* population. On the average, the derived *sh2* super-sweet corn populations flowered significantly later than the donor *sh2* population. The ASI, which is an indication of the extent of synchrony in flowering ranged from 1.5 to 3.5 days with a mean of 2.6 days. Eight of the derived *sh2* populations had significantly lower ASI than the donor *sh2* composite population.

In this study, orthogonal comparison revealed significant differences between the derived *sh2* populations and the donor *sh2* composite population for all the traits, with the derived *sh2* populations being superior to the donor *sh2* composite (Table 3).

DISCUSSION

Sweet corn cultivars are virtually non-existent in Nigeria, have narrow genetic base and poor adaptation to tropical environments. One strategy that could be adopted to broaden its genetic base, improve its adaptation and enhance its agronomic performance under tropical conditions is to cross with tropical field maize. In this study, some *sh2* super-sweet corn populations derived from crosses between *sh2* super-sweet corn population and tropical field maize genotypes were evaluated for their agronomic performance.

The significant effects of genotype and the higher contribution of the main effects of genotype to the total sum of squares observed in this study was a manifestation of the fact that the populations varied greatly in their performance. These genetic differences could be exploited in further breeding programmes. These significant genetic differences also indicated that the conversion of the field corn genotypes into super-sweet corn was effective in the development of new sweet corn populations different from the *sh2* donor parent. This was furthermore evidenced in the superiority of most of the derived populations over the *sh2* donor parent population for most of the traits studied. In this study, yield of marketable cobs averaged 6.84 t/ha. This was higher than the average yield of marketable fresh cobs of 5.03 t/ha reported by Kim et al. (2007) and 4.72

t/ha reported by Abe and Akinrinola (2015) for open pollinated varieties of normal endosperm tropical field maize. This observed yield potential reflects the great prospects that abound for sweet corn production in Nigeria. The findings of this study confirmed previous reports (Cartea et al., 1996; Malvar et al., 1997, 2001; Tracy, 2001; Butrón et al., 2008; Santos et al., 2014; Entringer et al., 2017) on the potential of tropical field maize genotypes in the improvement of sweet corn.

Conclusion

Significant genotypic differences were observed among the populations studied. Most of the derived *sh2* populations exhibited significantly superior performance relative to the *sh2* donor population for most of the traits. The results of the present study indicated that new super-sweet corn populations different from the donor parent could be developed by converting field corn genotypes into super-sweet corn through backcrossing and selection for the sweet corn trait. The observed genetic differences could be exploited in further breeding programmes. The superior populations could be further improved and used as base populations for inbred line development.

CONFLICT OF INTERESTS

The authors have not declared any conflict of interests.

ACKNOWLEDGEMENTS

The authors wish to thank Professor V.O. Adetimirin of the Department of Agronomy, University of Ibadan for generously providing the *sh2* donor population used in this study. The normal endosperm tropical field maize genotypes were sourced from the Maize Improvement Programme of IITA, Ibadan. This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

REFERENCES

- Abe A, Akinrinola TB (2015). Green ear yield potential of tropical field maize at two levels of nitrogen fertiliser application in Ibadan, Nigeria. *Ibadan Journal of Agricultural Research* 11(1):201-209.
- Adetimirin VO (2008). Stand establishment and early field vigour variation in a tropicalised shrunken-2 maize population. *Field Crops Research* 108:143-149.
- Brewbaker JL (1977). 'Hawaiian superweet #9' corn. *HortScience* 12:355-356.
- Butrón A, Álvarez A, Revilla P, Malvar RA, Rodríguez VM, Galarreta JIR, Ordás A (2008). Agronomic performance of sweetcorn populations derived from crosses between sweetcorn and field corn. *Journal of Agricultural Research* 6:378-384.
- Cartea ME, Malvar RA, Revilla P, Ordás A (1996). Improvement of early vigor and adaptation of sweet corn to European Atlantic coast with open-pollinated field corn populations. *Maydica* 41:119-125.

- Dewanto V, Wu X, Liu RH (2002). Processed sweet corn has higher antioxidant activity. *Journal of Agriculture and Food Chemistry* 50(14):4959-4964.
- Entringer GC, Vettorazzi JCF, Crevelari JA, Durães NNL, Catarina RS, Pereira MG (2017). Super-sweet corn breeding by backcross: A new choice for the Brazilian market. *Brazilian Journal of Agriculture* 92(1):12-26.
- FAO (2016). Food and Agriculture Organization of the United Nations statistical database, <http://www.faostat.fao.org/site/567/DesktopDefault.asp?PageID=567#ancor> (accessed on 26 August, 2017).
- Kim SK, Ajala SO (1996). Combining ability of tropical maize germplasm in West Africa II. Tropical vs. Temperate x Tropical origins. *Maydica* 41:135-141.
- Kim SK, Adetimirin VO, Yoon ST, Adepoju MA, Gbadamosi BA (2007). Green-maize potential of hybrid and open pollinated cultivars at varying levels of applied nitrogen: relationships with grain yield. *Tropical Science* 47(4):149-158.
- Kim SK, Efron Y, Khadr F, Mareck J, Fajemisin J (1987). Registration of 16 maize streak virus resistant tropical maize parental inbred lines. *Crop Science* 27:824-825.
- Lee SS, Yun SH, Kim JH (1999). Sugars, soluble solids, and flavour of sweet, super-sweet, and waxy corns during grain filling. *Korean Journal of Crop Science* 44(3):267-272.
- Lertrat K, Pulam T (2007). Breeding for increased sweetness in sweet corn. *International Journal of Plant Breeding* 1(1):27-30.
- Lizaso JI, Boote KJ, Cherr CM, Scholberg JMS, Casanova JJ, Judge J, Jones JW, Hoogenboom G (2007). Developing a Sweet Corn Simulation Model to Predict Fresh Market Yield and Quality of Ears. *Journal of American Society of Horticultural Science* 132(3):415-422.
- Malvar RA, Cartea ME, Revilla P, Butrón A, Ordás A (2001). Checking performance of field corn inbreds as donors of favourable alleles to improve early vigor and adaptation of sweet corn hybrids to European conditions. *Maydica* 46:187-193.
- Malvar RA, Revilla P, Cartea ME, Ordás A (1997). Field corn inbreds to improve sweet corn hybrids for early vigor and adaptation to European conditions. *Maydica* 42:247-255.
- Ogunbodede BA (1999). Green maize production in Nigeria in the new millennium- Prospects and Problems. *Genetics and food security in Nigeria in the twenty-first century*. Olaoye, G. and Ladipo, D.O. Eds. Genetics Society of Nigeria special publication pp. 33-37.
- Osayintola OJ, Mareck JH, Akingbala JO (1992). Effect of time of harvest and variety on the quality of boiled green field maize (*Zea mays* L.). *Tropical Science* 32:369-376.
- Qi X, Zhao Y, Jiang L, Cui Y, Wang Y, Liu B (2009). QTL analysis of kernel soluble sugar content in super-sweet corn. *African Journal of Biotechnology* 8:6913-6917.
- Santos PHAD, Pereira MG, Trindade RS, Silva da Cunha K, Entringer GC, Vettorazzi JCF (2014). Agronomic performance of super-sweet corn genotypes in the north of Rio de Janeiro *Crop Breeding and Applied Biotechnology* 14:8-14.
- SAS Institute Inc. (2003). SAS/STAT user's guide, version 9.1.3. SAS Institute Inc., Cary, NC, USA.
- Teixeira FF, Miranda RA, Paes MCD, Souza SM, Gama EEG (2013). *Melhoramento do milho doce*. Sete Lagoas: Embrapa Milho e Sorgo. 32 p
- Tracy WF (1997). History, genetics, and breeding of super-sweet (*shrunk2*) sweet corn. *Plant Breeding Reviews* 14:189-236.
- Tracy WF (2001). Sweet corn. *Speciality corns*. Hallauer, A.R. Ed. 2nd ed. CRC Press, Boca Raton, Florida. pp. 155-197.
- Tracy WF, Whitt SR, Buckler ES (2006). Recurrent mutation and genome evolution: example of Sugary1 and the origin of sweet maize. *Crop Science* 46:1-7.
- U.S. Department of Agriculture (USDA) (2017). Sweet corn data. https://www.nass.usda.gov/Statistics_by_Subject/result.php?49AD4F93-5D68-3E98-BA30-EE901B111899§or=CROPS&group=VEGETABLES&comm=SW EET%20CORN (Accessed 24 December, 2018).
- Yousef GG, Juvik JA (2002). Enhancement of seedling emergence in sweet corn by marker-assisted backcrossing of beneficial QTL. *Crop Science* 42(1):96-104.
- Zan GH, Brewbaker JL (1999). Seed quality of isogenic endosperm mutants in sweet corn. *Maydica* 44:271-277.